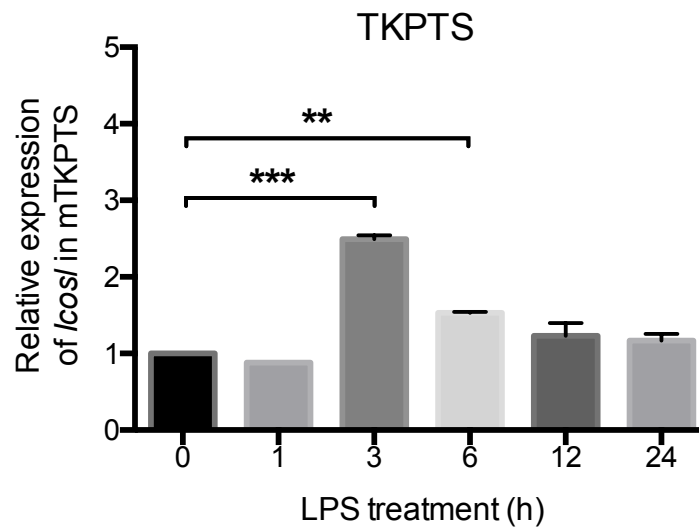




Supplemental Figure 1.

Expression levels of *Icosl* are increased by LPS in mouse kidney proximal tubule cell line (TKPTS). Relative mRNA expression measured by quantitative PCR targeting *Icosl* in mouse TKPTS cells that were treated with 50 µg/ml LPS for 1, 3, 6, 12 and 24 hours. QPCR analysis of *Icosl* mRNA was normalized to the expression of *Gapdh* and presented relative to *Icosl* expression levels in untreated cells. $n=3$ biologically independent samples. Data are mean $\pm$ SD; ** $P<0.01$, *** $P<0.001$; One-way ANOVA with Tukey's multiple comparison test.

A

1. B7.1; NP_005182.1 T-lymphocyte activation antigen CD80 precursor, B7.1
2. B7.2; NP_787058.4 T-lymphocyte activation antigen CD86 isoform 1 precursor, B7.2
3. ICOSL; NP_056074.1 ICOS ligand isoform a precursor
4. PDL-1; NP_054862.1 programmed cell death 1 ligand 1 isoform a precursor

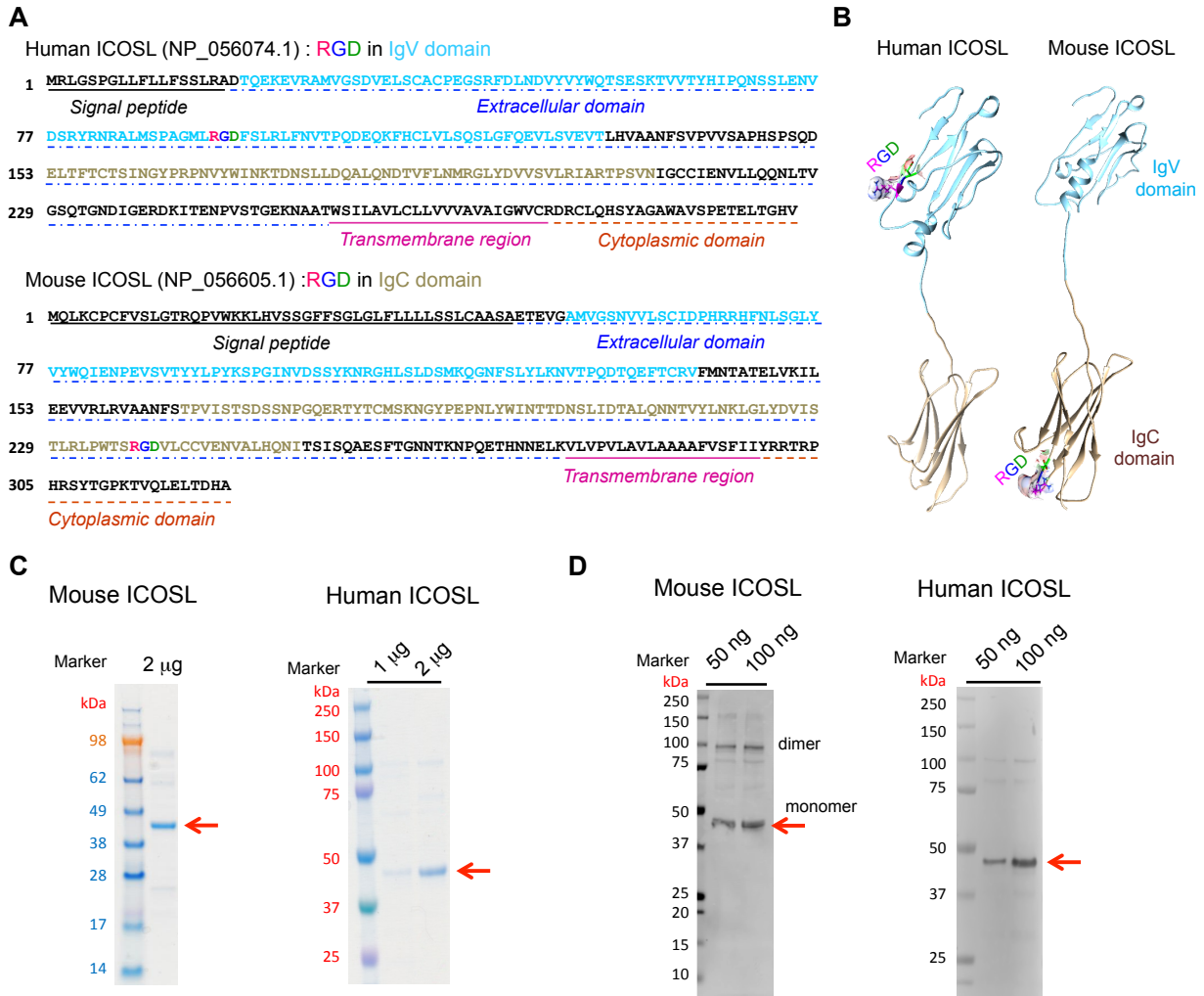
1.	NP_005182.1	MGHTRRQGTSPSKCPY----LNFFQLLVLAGLSHFCSGVIHVTKEVKEVATLSCGHNV-S	55
2.	NP_787058.4	-----MDPQCTMGLSNILFVMAFLLSGA-----APLKIQAYFNETADLPQCFANSQ	46
3.	NP_056074.1	-----MRLGSPGLLFLFFSSLRA----DTQEKEVRAMVGSDELSCACPEGS	43
4.	NP_054862.1	-----MRIF-AVFIFMTYWHLLNAFTVTVPKDLVVEYGSNMTIECKFPVEK	46
		: * . * . : *	
1.	NP_005182.1	VEELAQTRIYWQKEKMMV-LTMMSGDM----NIWPEYKNRTIFDI----TNNLSIVILAL	106
2.	NP_787058.4	NQSLSELVVFWDQENLV-LNEVYLGEKEFDSVHSKYMGRTSFDS----D-SWTLRLHNL	100
3.	NP_056074.1	RFDLNDVYVWQTSESKTVVYTHIPQNSSLENVDSTRYRNALMSPAGMLR ^{GD} FSLRLENV	103
4.	NP_054862.1	QLDLAALIVWEMEDKNI-IQ-FVHGEEDLKVQHSSYRQARLLKDLQSLGNAALQITDV	104
		. * : : * : . : : :	
1.	NP_005182.1	RPSDEGTYECVVLKYEKDAFKREHLAEVTLVKADFPTPSISDFEIPT-SNIRRIICSTS	165
2.	NP_787058.4	QIKDKGLYQCIHHKKPTGMIRIHQMNSLSVLANFSQPEIVPISNITENVYINLTCSSI	160
3.	NP_056074.1	TPQDEQKFHCLVLSQ-SLGFQEVLSVEVTLHVAANFSVPVVSAPHSPS-QDELTFCTCTSI	161
4.	NP_054862.1	KLQDAGVYRCMISYG-GADYKRIT-----VKVNAPYNKINQRIILVVDPTSEHELTQ-A	157
		. * : : * : . : * * : : *	
1.	NP_005182.1	GGFPEPHLSWLENGEELNAI----NTTVSQDPETELYAVSSKLDLFNM---TTNHSFMCLI	218
2.	NP_787058.4	HGYPEPKKMSVLLRTKNSTIEYDGIMQKSQDNVTELYDVSISLSVSFPDVTSNMTIFCIL	220
3.	NP_056074.1	NGYPRENVYWINKTDN-SLLDQALQNDTVFLNMRGLYDVVSVLRIAR---TPSVNIGCCI	217
4.	NP_054862.1	EGYPKAEVIWTSSDHQVL---SGKTTTNSKREEKLFNVTSTLRINT---TTNEIFYCTF	211
		* : * . : : * : * * * . * . : * :	
1.	NP_005182.1	KYGHLRVNQ-----T-----FNWNTTKQEHFPDNLPSWAITLISVNGIFVICCLTYCF	267
2.	NP_787058.4	ETDKTRLLS-----SP-----FSIELEDPPQPPD--HIPWITAV---LPTVIICVMVFCL	265
3.	NP_056074.1	ENVLLQQNLTVGSQTGNDIGERDKITENPVSTGEKNAATWSILA-----VLCLLVVVA	270
4.	NP_054862.1	RRLDPEENHTAE-----LVIPELPLAH-----PPNERTHLVILGAILLCLGV-AL	255
		. : : : : : : : : : : *	
1.	NP_005182.1	APRCRERRRNE-----RLRRESVRPV-----	288
2.	NP_787058.4	ILWKWKKKRPRNSYKCGTNTMERESEQTKKREKIHIPERSDEAQRVFKSSKTSSCDKS	325
3.	NP_056074.1	VAIGWVCRDRCLQHSYAGAVSPETE-LTG-----HV-----	302
4.	NP_054862.1	TFIFRLRKGRMMDVKKCIQDNTNSKKQS-----DTHLEET-----	290
		: . : :	
1.	NP_005182.1	----	288
2.	NP_787058.4	DTCF	329
3.	NP_056074.1	----	302
4.	NP_054862.1	----	290

B

		Protein Accession No.	Organism	Gene
1. NP_056074.1			H.sapiens	ICOSLG
2. XP_006723963.1			H.sapiens	LOC102723996
3. XP_003319157.1			P.troglodytes	ICOSLG
4. XP_002803158.1			M.mulatta	LOC722191
5. XP_005639015.1			C.lupus	ICOSLG
6. XP_005202100.1			B.taurus	ICOSLG
7. NP_056605.1			M.musculus	Icosl
8. XP_006256322.1			R.norvegicus	Icoslg
9. NP_990415.1			G.gallus	ICOSLG
1. NP_056074.1	1	MRL-----GSPGLLFLFSSLRADTQEKEVRAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--	68	
2. XP_006723963.1	1	MRL-----GSPGLLFLFSSLRADTQEKEVRAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--	68	
3. XP_003319157.1	1	MRL-----GSPGLLFLFSSLRADTQEKEVRAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--	68	
4. XP_002803158.1	1	MRL-----GSPGLLFLFSSLRADTQEKEVRAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--	68	
5. XP_005639015.1	1	MWL-----RCPGVLLLLGLTQANTQEVEKALVGSSVELRCVFPERHTFDLDDLYVYWQISVVVGQ---PKTVTYTYS--	70	
6. XP_005202100.1	1	MRP-----RSPGLFLLLLWGLQAESQE-EVRAMVGSDELRCIYPEKNSFDLNDLYVYWQISMAGKGNVDSVVTYHLS--	72	
7. NP_056605.1	1	MQLKPCFVSLGTRQCPVWKHLHVSSGFFSGLGLFLLSSSLCAASAEVKGAMVGSNNVLSCLDPHRRHFNLSGLYVYWQI-ENP-----EVSVTYYLP--	93	
8. XP_006256322.1	1	MQL-----NRFFSGPLGLFLLFCSLCEAEVKEVNAMVGSDELRCVYPRRSHFSLDDLYVYWQIVDEA-----KTVVTYYTLP--	75	
9. NP_990415.1	1	MKR-----LGYGFLLLFLHLILRAVTALEKTIISKPGDNATLSCYIYANRG-FDLSLRVYWQIDGVEGSKSCSVVHALIS--	72	
1. NP_056074.1	26	RAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--QNSSLENVDSRYNRNALMS PAGMLRGDFSRLRNFVTPQDEQKFHCLVL-S	117	
2. XP_006723963.1	26	RAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--QNSSLENVDSRYNRNALMS PAGMLRGDFSRLRNFVTPQDEQKFHCLVL-S	117	
3. XP_003319157.1	26	RAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--QNSSLENVDSRYNRNALMS PAGMRRGDFSRLRNFVTPQDEQKFHCLVL-S	117	
4. XP_002803158.1	26	RAMVGSDELSCACPEGSFRDLNDVYVYWTSES-----KTVVTYHIP--QNSSLDNVDSHYNRNALMS PAGMQRGDFSRLRNFVTPQDEQKFHCLVL-S	117	
5. XP_005639015.1	26	KALVGSSVELRCVFPERHTFDLDDLYVYWQISVVVGQ---PKTVTYTYS--GNSSTGQEDDRYDRARLS PESMQRGDFSRLHLYNITPYDEQKFHCLV-F-R	119	
6. XP_005202100.1	25	RAMVGSDELRCIYPEKNSFDLNDLYVYWQISMAGKGNVDSVVTYHLS--GNSASGHGDNHYKDRARLSLDSMRQGFDSLHLRNVTTPQDEQKFHCLV-F-R	121	
7. NP_056605.1	51	GAMVGSNNVLSCLDPHRRHFNLSGLYVYWQI-ENP-----EVSVTYYLP--YKSPGINVDSSYKNGHLSLDSMRQGNFSLYLNKVTTPQDTQEFTCRVFMN	143	
8. XP_006256322.1	30	NAMVGSDELRCVYPRRSHFSLDDLYVYWQIVDEA-----KTVVTYTLP--SANESSSTIHVSNSYKNRAHLSPLMKRGDFSRLHLYNVTTPQDTQEFTCLV-F-R	124	
9. NP_990415.1	26	ISKPGDNATLSCYIYANRG-FDLSLRVYWQIDGVEGSKSCSVVHALIS--GQDNESQQCSQFKNRTQLLWDKLGDDGDFSLLLNVNRQSDHEITYKCVVM--	120	
1. NP_056074.1	118	QSLGFQEVLSVEVTLHVAANFSPVVSAP--HSPSQD-ELTFTCTCSINGYPRPNVYWINKTDSNLLDQALQNDTVFLNMRGLYDVVSVLRIARTPSVNI	213	
2. XP_006723963.1	118	QSLGFQEVLSVEVTLHVAANFSPVVSAP--HSPSQD-ELTFTCTCSINGYPRPNVYWINKTDSNLLDQALQNDTVFLNMRGLYDVVSVLRIARTPSVNI	213	
3. XP_003319157.1	118	QSLGFQEVLSIEVTLHVAANFSPVVSAP--HSPSQD-ELTFTCTCSINGYPRPNVYWINKTDSNLLDQALQNDTVFLNTRGLYDVVSVLRIARTPSVNI	213	
4. XP_002803158.1	118	QSLGFQEVLSVVVTLHVAANFSPVVSAP--HNPSQD-KLTFTCTCSINGYPRPNVYWINKTDSNLLDQALQNDTVFLNTRGLYDVVSVLRIARTPSLNI	213	
5. XP_005639015.1	120	KSLELKKIL-DVEVTLHVAANYSMFVVSAP--SDPSKDELFTFTCTSVNGYPRPNVYWINKTDSNVLDDGGLQNHTVSVNAQGLYDVVSVLRVFPWAPSINV	216	
6. XP_005202100.1	122	KSLELKKIL-EVAVTLHVAANYSMFVVSAP--PSQDEELFTFTCTSVNGYPRPNVYWINKTDSNVLDSALQNNTVSVNARGLYDVVSVLRIGRTPHVN	215	
7. NP_056605.1	144	TATELVKIL-EEVRLRVAANFSTFVISTS--DSSNPGQERTYTCMSKNGYEPENLYWINTTDSNLDLIDTALQNNTVYLNKGLYDVISTLRFPWTSGRGDV	240	
8. XP_006256322.1	125	MSTVLGKAL-EEVRLRVAANFSTFVISTS--GSSDPGQERTYTCMSKNGYEPENLYWINTTDLIDTALQNNTVYLNELGLYDVVSTLRFPWTHVDV	221	
9. NP_990415.1	121	QTIEYTRVHQEQVVLSSLAASYSPILSGPIRNSYSTGEEVTFSCRSNDNGYEPENLYWINKTDNTRLSQS--DFNITQHPDGTYSVLSTLKNVATSDMQ	218	
1. NP_056074.1	214	GCCIEENVLLQQLNTVGSQTGNIDIGERDKITENPV--STGEKNAATWSILAVLCLLVVVAIGA-WVCRDRCL-----	282	
2. XP_006723963.1	214	GCCIEENVLLQQLNTVGSQTGNIDIGERDKITENPV--STGEKNAATWSILAVLCLLVVVAIGA-WVCRDRCL-----	282	
3. XP_003319157.1	214	GCCIEENVLLQQLNTVGSQTGNIDIGERDKITENPV--STGEKNAATWSILAVLCLLVVVAIGA-WVCRDRCL-----	282	
4. XP_002803158.1	214	GCCIEENVLLRQNLTVGSQTGNIDIGERDKITENPV--STREKTEVTVSWLAVLCLLVVVAIGA-WVCRDRWL-----	282	
5. XP_005639015.1	217	GCCIEENVLLHQNLTNR-PAEPIPGTEPGNTGSHRD-VHRRDHGAVLSAVAVLLVLAVLGA-VG-CVCRSRCPRTDPCRLRLKTRRLGRGQGFCEMPGG	313	
6. XP_005202100.1	216	GCCIEENVLLHQNLTSG-QTETFTGTDKSDITEDPVDGSPSGKHQGVFSLAVLAVIIVLALATG-WLCRSRCP-----	285	
7. NP_056605.1	241	LCCVENVALHQNITSISQAESFTGN-N--TKNPQE-THNNELKVLFPVLAVLAAAFVSFII--YR-RTR-P-----	304	
8. XP_006256322.1	222	ICCVENVALHQNITSISRADSFTGSMN--TERPQE-IHREATKVLFFVLAALLAVVVVIFIIVLYRCRRR-P-----	289	
9. NP_990415.1	219	ECFTENKVLQENTSANYTEEMQNGSS--TGSHKDAAGGGQGAQAAVSVVILMAFLTVLIC-WLWRRRSF-----	287	
1. NP_056074.1	283	-----QHSYAGAWA-----VSPETELTGHV-----	302	
2. XP_006723963.1	283	-----QHSYAGAWA-----VSPETELTGHV-----	302	
3. XP_003319157.1	283	-----QHSYAGAWA-----VSPETELTGHV-----	302	
4. XP_002803158.1	283	-----RRSYAGVWA-----VRPEPELAGEFAMGKQVQALAPTPTGCGSEQVQALAPTPTGCSQQVQALAPTPT	343	
5. XP_005639015.1	314	TADGSLRIDPGRPGGQGGPAAPVGAVKKGASWPRDLSLEGTRRIVLDAVASVPAAPILG-----	374	
6. XP_005202100.1	286	-----PKSQTGAQV-----ARPKLALAEHV-----	305	
7. NP_056605.1	305	-----HRSYTGPKT-----VQLELTDHA-----	322	
8. XP_006256322.1	290	-----CQSYTGPR-----VQLELTGECTGERPVG-----	315	
9. NP_990415.1		-----QLVSYTAPV-----	296	
1. NP_056074.1		-----		
2. XP_006723963.1		-----		
3. XP_003319157.1		-----		
4. XP_002803158.1	344	TGCSEQVQALQVQALAPTPTGCGSEQVQALVPSPHGVFRGGPGTCAHPYQDVRSRHRCLLSAAATPAALGRPLSPPEKTDVCLQNGKGCFFMLSLHFP	443	
5. XP_005639015.1		-----		
6. XP_005202100.1		-----		
7. NP_056605.1		-----		
8. XP_006256322.1		-----		
9. NP_990415.1		-----		
1. NP_056074.1		-----		
2. XP_006723963.1		-----		
3. XP_003319157.1		-----		
4. XP_002803158.1	444	KAMFDRSSPPRAWTGLP	460	
5. XP_005639015.1		-----		
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9. NP_990415.1		-----		

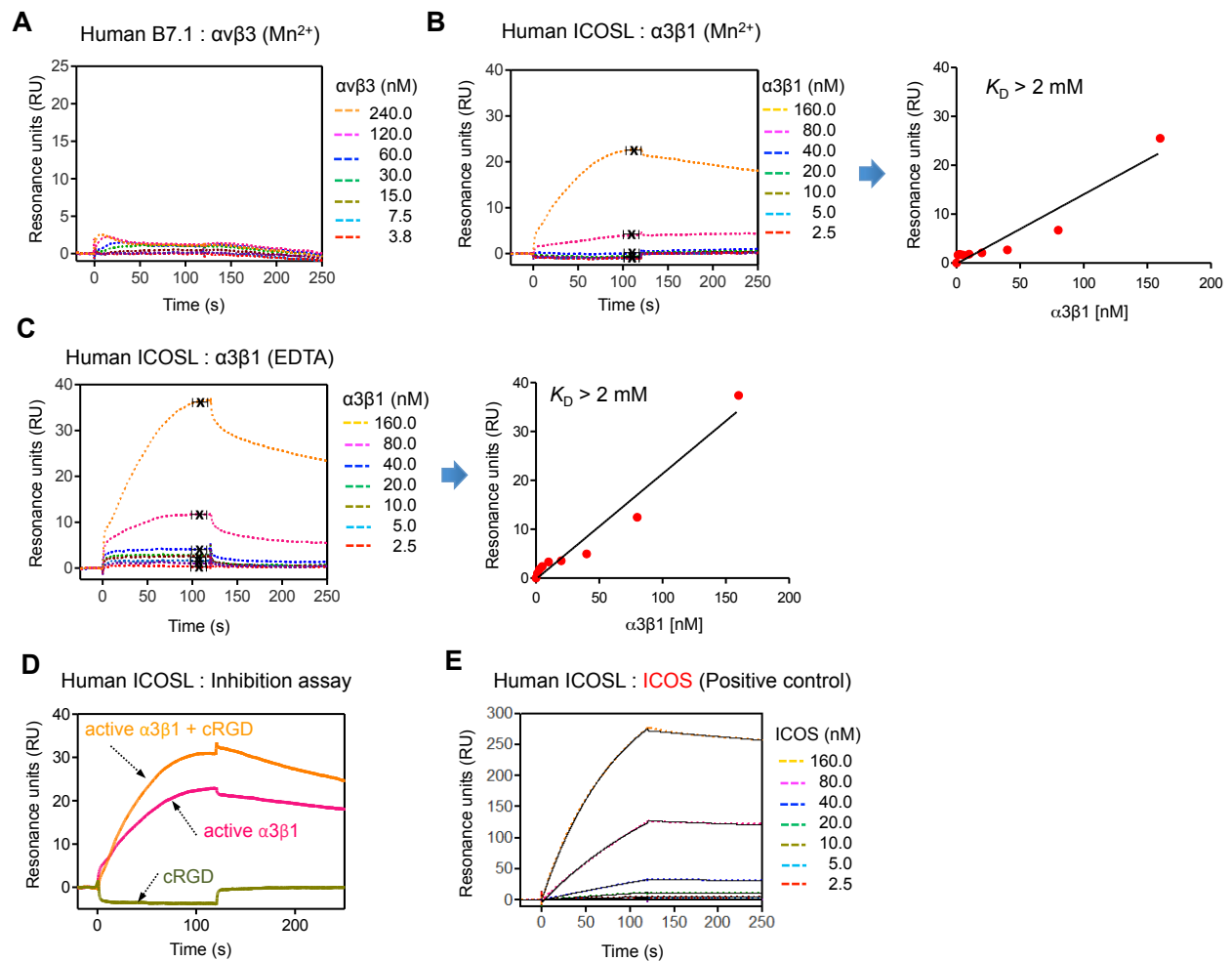
Supplemental Figure 2.

Among B7 family members, only ICOSL gene contains an RGD motif which is highly conserved across different species. (A) Multiple sequence alignments of human B7 families; B7.1, B7.2, ICOSL and PDL-1 were aligned using the Clustal Omega server (1). (B) The ICOSL proteins used in this alignment were derived from HomoloGene database available at NCBI (<http://www.ncbi.nlm.nih.gov/homologene/>) and were 1. NP_056074.1 (H.sapiens), 2. XP_006723963.1 (H.sapiens), 3. XP_003319157.1 (P.troglodytes), 4. XP_002803158.1 (M.mulatta), 5. XP_005639015.1 (C.lupus), 6. XP_005202100.1 (B.taurus), 7. NP_056605.1 (M.musculus), 8. XP_006256322.1 (R.norvegicus), and 9. NP_990415.1 (G.gallus). In evolutionarily advanced vertebrates such as human (H.sapiens), chimpanzee (P.troglodytes), monkey (M.mulatta), and wolf (C.lupus), the RGD sequences are located in the IgV domain. In contrast, mouse (M.musculus) ICOSL has its RGD domain in the IgC domain. The RGD sequences are highlighted in red.



Supplemental Figure 3.

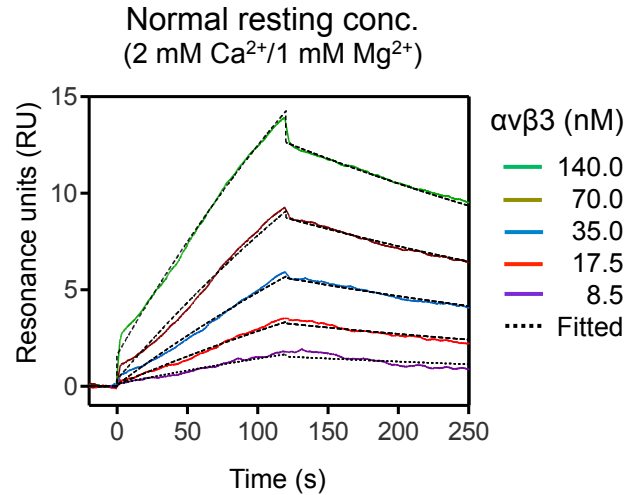
Both human and mouse ICOSL contain an RGD motif at an exposed loop region. (A) Sequence features of the RGD motif in human and mouse ICOSL domains. **(B)** Ribbon diagram of a three-dimensional model of the extracellular domains of human (left) and mouse ICOSL (right). **(C, D)** Quality and purity of the purified mouse and human recombinant ICOSL proteins were assessed using Coomassie Blue staining **(C)** and Western blotting **(D)**. In SDS-PAGE gel staining, 1 or 2 μ g of recombinant proteins were loaded in lanes 1 and 2, respectively. For Western-blotting, 50 and 100 ng of recombinant protein were separated on SDS-PAGE, transferred to a nitrocellulose membrane and incubated with rabbit anti-ICOSL antibody (MyBiosource, 1:500).



Supplemental Figure 4.

ICOSL shows no appreciable binding to $\alpha 3\beta 1$ integrin. (A) SPR sensorgrams depicting interaction between immobilized human B7.1 and active (Mn^{2+} -induced) $\alpha\beta 3$ integrin at various concentrations (0-240 nM). (B, C) SPR sensorgrams depicting interaction between immobilized human ICOSL and active (B, 2 mM Mn^{2+} in binding buffer) or inactive (C, 3 μ M EDTA in binding buffer) $\alpha 3\beta 1$ integrin. On the *right-hand side* of each sensorgrams are the steady-state affinity fitting curves of $\alpha 3\beta 1$ integrin (0-160 nM) binding to ICOSL. The K_D values determined for each of active or inactive $\alpha 3\beta 1$ integrin were > 2 mM. (D) SPR illustrating the inhibition of $\alpha 3\beta 1$ binding to ICOSL by the synthetic cRGD peptide. Injection of active $\alpha 3\beta 1$ (160 nM) resulted in the binding signal for immobilized human ICOSL (pink curve). Preincubation with cRGDfv (30 μ M) did not inhibit interaction with ICOSL, indicating that the RGD peptide did not compete with ICOSL for binding to $\alpha 3\beta 1$ (orange curve). The cRGDfv alone was used as a

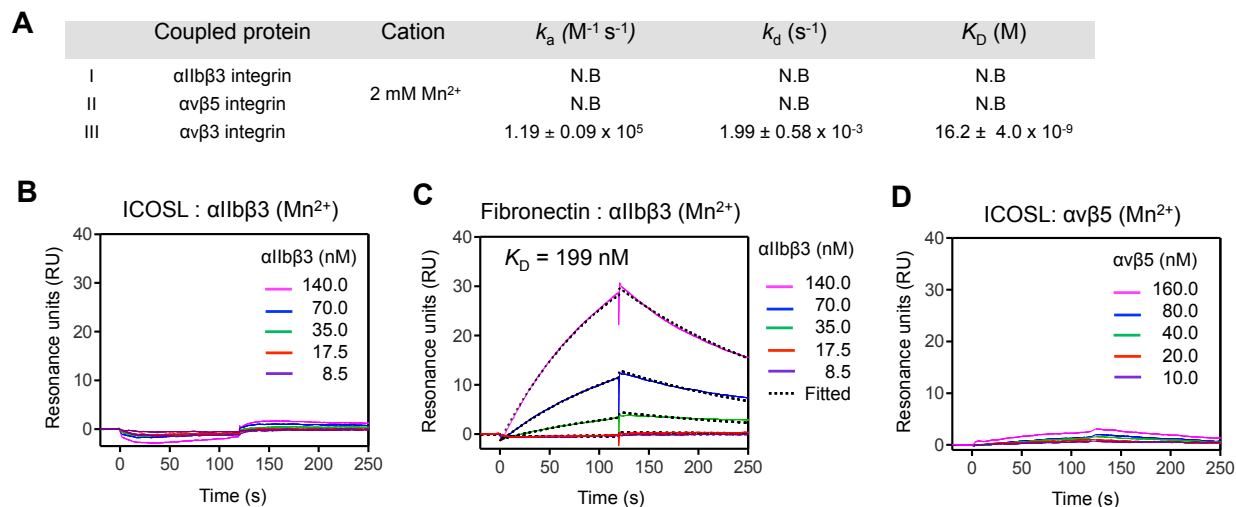
control (green curve). (E) SPR sensorgrams depicting interaction between immobilized human ICOSL and ICOS at various concentrations (0-160 nM). Kinetic constants (k_a , k_d) were derived from locally fitting the association and dissociation phases using a 1:1 Langmuir binding model (black fitting curves) and affinities were calculated as $K_D=k_d/k_a$. The average K_D values were determined from at least three independent experiments (A-E).



Concentrations of Ca ²⁺ and Mg ²⁺	k_a (M ⁻¹ s ⁻¹)	k_d (s ⁻¹)	K_D (M)
2 mM, 1 mM	$5.61 \pm 0.97 \times 10^3$	$2.18 \pm 0.14 \times 10^{-3}$	$400 \pm 40 \times 10^{-9}$
0.2 mM, 0.1 mM (Figure 2H)	$1.51 \pm 0.39 \times 10^5$	$3.26 \pm 0.10 \times 10^{-3}$	$21.3 \pm 1.2 \times 10^{-9}$

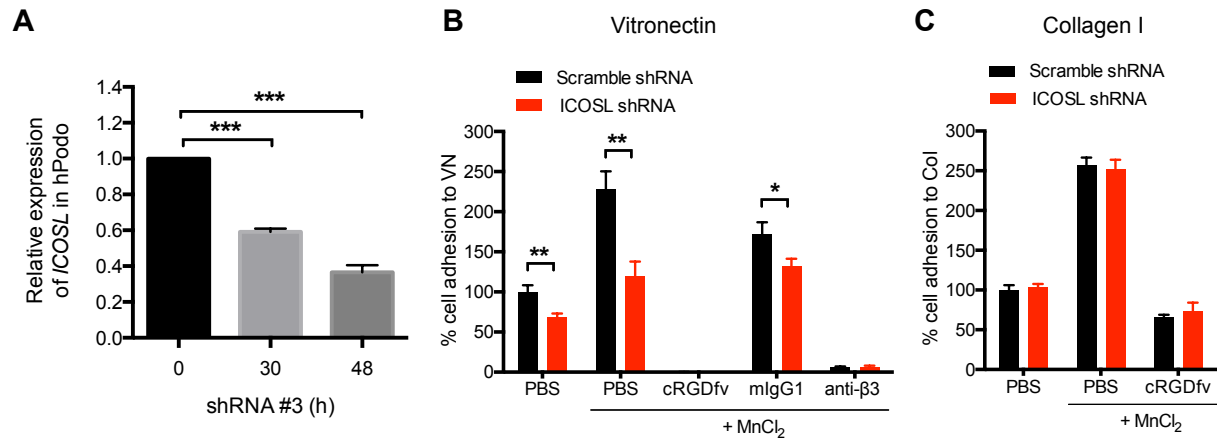
Supplemental Figure 5.

The binding between ICOSL and αvβ3 integrin is inhibited under normal resting serum concentrations of Ca²⁺ and Mg²⁺ (2 mM and 1 mM, respectively). *Top:* Representative SPR sensorgram showing the binding between mICOSL protein and αvβ3 integrin under normal resting serum concentrations of Ca²⁺ and Mg²⁺ (2 mM and 1 mM, respectively). *Bottom:* Table summarizing the binding characteristics of ICOSL protein towards αvβ3 integrin under different concentrations of Ca²⁺ and Mg²⁺ (normal vs low). The average K_D values were determined from three independent experiments. Kinetic constants (k_a , k_d) were derived from locally fitting the association and dissociation phases using a 1:1 Langmuir binding model (black fitting curves) and affinities were calculated as $K_D = k_d/k_a$.



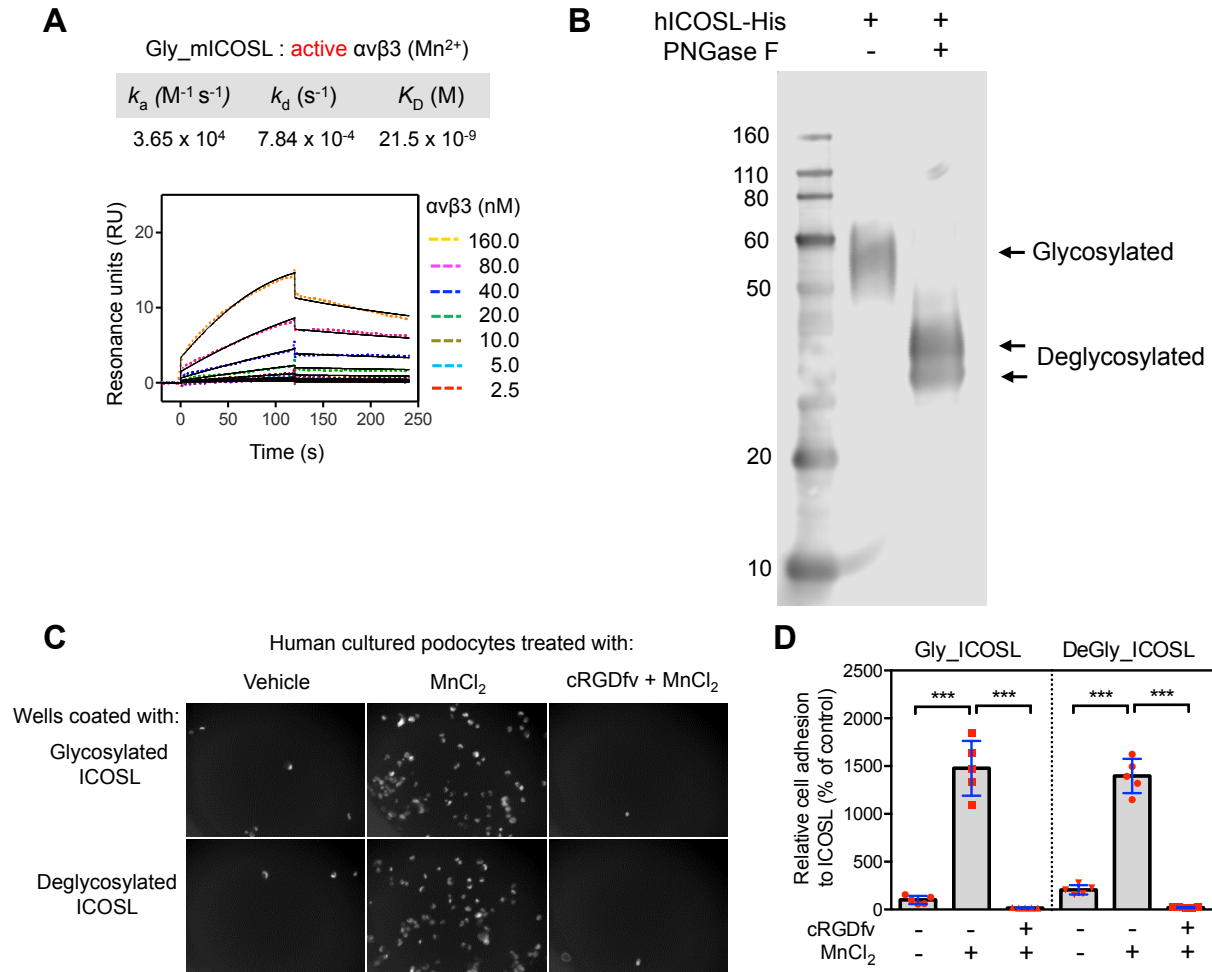
Supplemental Figure 6.

ICOSL selectively binds to $\alpha v \beta 3$ but not to $\alpha I I b \beta 3$ or $\alpha v \beta 5$. (A) Table showing the binding characteristics of human ICOSL protein towards three different RGD-dependent integrins. N.B, no binding. (B-D) Representative sensorgrams depicting interaction of $\alpha I I b \beta 3$ integrin with ICOSL (B) or fibronectin (C; positive control), and between $\alpha v \beta 5$ and ICOSL (D) at various concentrations of integrins (0-140 or 0-160 nM in running buffer containing 2 mM Mn^{2+}). The average K_D values were determined from three independent experiments. Kinetic constants (k_a , k_d) were derived from locally fitting the association and dissociation phases using a 1:1 Langmuir binding model (black fitting curves) and affinities were calculated as $K_D = k_d/k_a$.



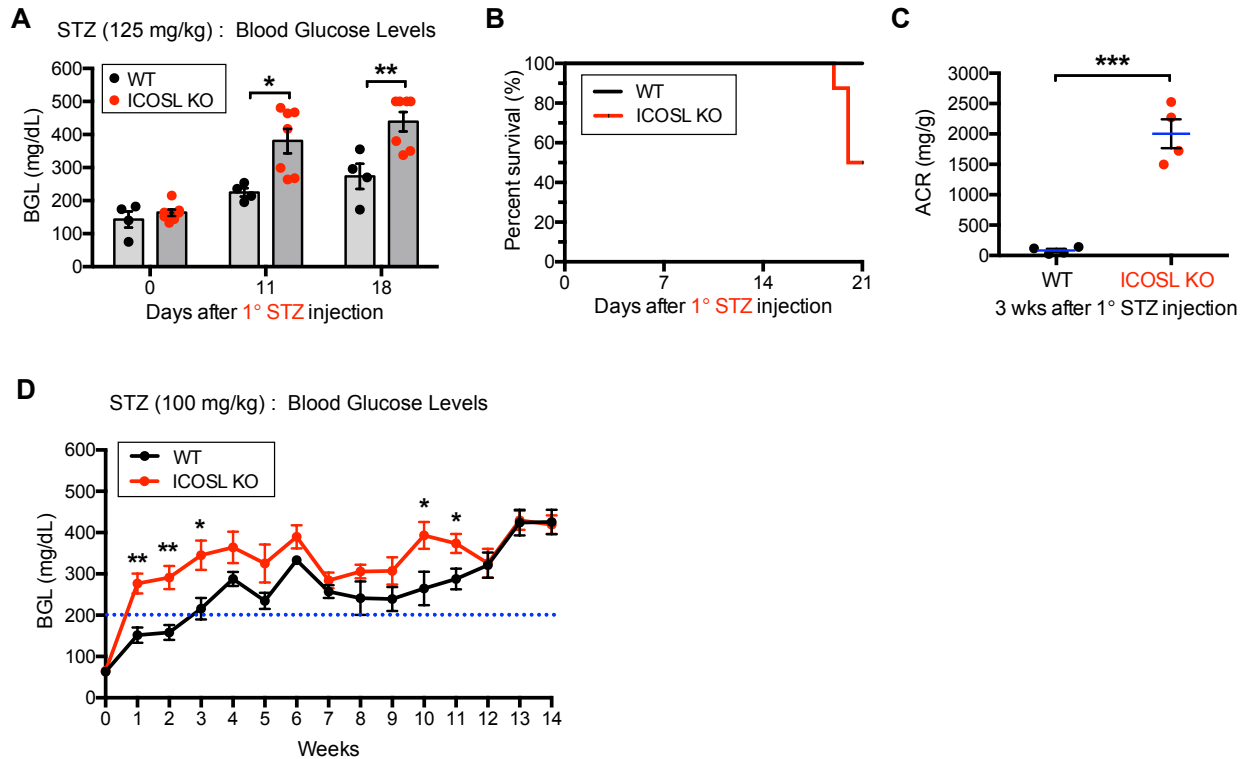
Supplemental Figure 7.

Knockdown of ICOSL inhibits $\beta 3$ integrin-dependent podocyte adhesion. (A) *ICOSL* mRNA expression is reduced in human podocytes after treatment with shRNA-expressing lentivirus. Human podocyte cell lines were infected with a shRNA-scrambled control or shRNA-*ICOSL* #3 (selected among 4 shRNAs targeting *ICOSL*) lenti-virus. Twenty four hours after transduction, the cells were replaced with regular growth media and incubated for 30 to 48 hours. The level of *ICOSL* mRNA was determined by qPCR. Data are normalized to *GAPDH* expression ($n=3$ biological replicates). Data are mean $\pm$ SD; *** $P<0.001$; One-way ANOVA with Tukey's multiple comparison test. (B and C) Loss of ICOSL function on human cultured podocytes measured by cell adhesion assay. Red bars represent adhesion levels of cells that were treated with shRNA targeting *ICOSL* while black bars represent levels from cells treated with a scrambled shRNA. Quantification of cell adhesion shows significantly reduced levels after treatment with shRNA targeting *ICOSL* on vitronectin-coated plates. The involvement of ICOSL was observed in the binding of cells in an RGD-dependent integrin manner when cells were plated on vitronectin (B), but not when plated on collagen I (C). Controls for this assay included the use of a competing peptide (cRGDfv) and an antibody directed against $\beta 3$ integrin (or mIgG₁ as an isotype control) in combination with an integrin activator, MnCl₂. Data are shown as mean $\pm$ SEM and from three independent experiments; * $P<0.05$, ** $P<0.01$; One-way ANOVA with Tukey's multiple comparison test.



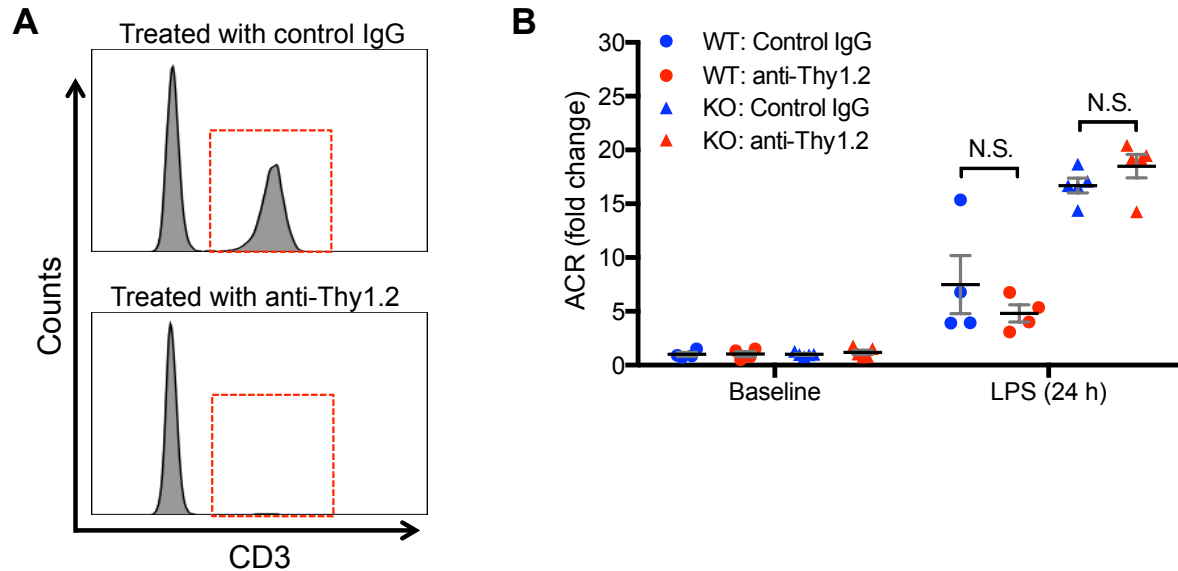
Supplemental Figure 8.

Glycosylation of ICOSL does not affect its ability to bind active $\alpha\beta 3$ integrin. (A) SPR kinetics analysis determined the equilibrium dissociation constant ($K_D=21.5$ nM) between immobilized glycosylated mouse ICOSL and $\alpha\beta 3$ integrin at various concentrations (0-160 nM). Kinetic constants (k_a , k_d) were determined by using a 1:1 Langmuir binding model (black fitting curves) and affinities were calculated as $K_D=k_d/k_a$. (B) PNGaseF treatment was performed following the manufacturer's protocol to deglycosylate human ICOSL-His protein (Thermo Scientific), followed by immunoblot analysis of the glycosylated and deglycosylated forms of the ICOSL protein using anti-His antibodies. Protein bands were visualized by the Odyssey imaging system. (C) Adhesion assay using cultured human podocytes. (D) Quantification of these adhesion assays (see Methods for details on the assay and how quantification was carried out). Data are mean $\pm$ SD and from two independent experiments; * $P<0.05$, ** $P<0.01$, *** $P<0.001$; One-way ANOVA with Tukey's multiple comparison test (D).



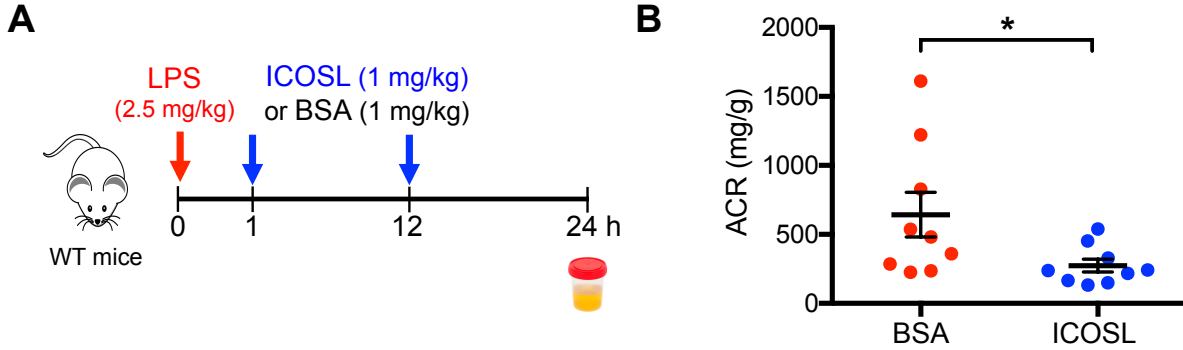
Supplemental Figure 9.

ICOSL deficient mice are more susceptible to the toxicity of STZ (125 mg/kg). (A-C) Diabetic ICOSL KO mice rapidly developed higher levels of hyperglycemia as compared to the WT mice ($n=4-8$ per group). (A) Eleven and 18 days after STZ treatment (125 mg/kg), the levels of hyperglycemia in ICOSL KO were significantly increased. (B) Survival rate was monitored for 21 days post STZ injection. (C) Albumin-to-creatinine ratio (ACR, mg/g) was calculated for surviving STZ-injected mice 21 days post injection. (D) Both BALB/c WT (black dots) and ICOSL KO (red dots) mice developed hyperglycemia by low dose STZ injection (100 mg/kg) ($n=5-6$ per group). The diabetic ICOSL KO mice rapidly developed higher levels of hyperglycemia as compared to WT mice. Three weeks after STZ treatment, the levels of hyperglycemia in either strain were sustained until 14 weeks. Data are mean $\pm$ SEM; * $P<0.05$, ** $P<0.01$, *** $P<0.001$; Multiple unpaired t test with the Holm-Sidak comparisons test (A, D); Student's two-tailed, unpaired t test (C).



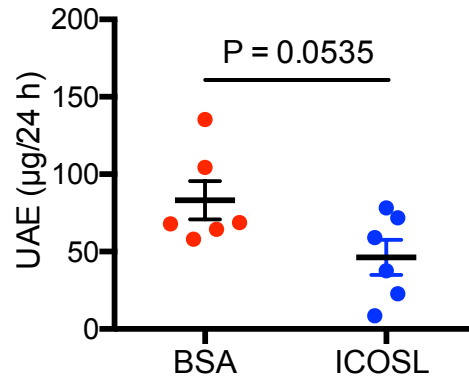
Supplemental Figure 10.

The renoprotective effect of ICOSL is not likely ICOS-dependent. Both WT and ICOSL KO mice were intravenously injected with monoclonal antibody against Thy1.2 (5 mg/kg) or rat IgG2a (5 mg/kg) 24 hours prior to LPS injection (2.5 mg/kg, i.p) (n=4 for WT+control IgG and WT+anti-Thy1.2, n=5 for KO+control IgG and KO+anti-Thy1.2). Urine and blood samples were collected at baseline and 24 hours after LPS treatment. Blood leukocytes were labeled with fluorescently conjugated antibodies specific for mouse CD3. **(A)** Representative histogram of flow cytometric analysis showing T cell depletion in mice treated with anti-mouse Thy1.2. **(B)** Albumin/creatinine (ACR) levels in WT and ICOSL KO mice treated with either control IgG or anti-Thy1.2. ACR levels normalized to baseline are shown as fold change. There was no statistically significant difference in ACR levels between T cell-depleted and control groups in both WT and KO mice. Data are mean $\pm$ SEM; N.S., not significant; Two-way ANOVA with Tukey's multiple comparison test.



Supplemental Figure 11.

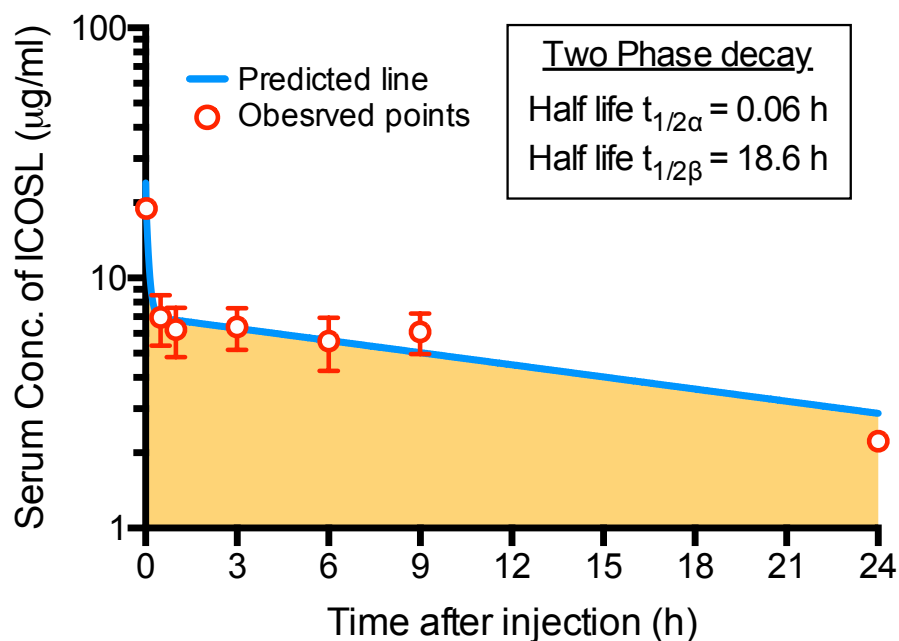
Administration of ICOSL WT reduces proteinuria in LPS-injected WT mice. BALB/c WT mice were challenged with 2.5 mg/kg LPS, then intravenously injected with mouse ICOSL protein (1 mg/kg) or BSA (1 mg/kg) as a protein control 1 and 12 hours after LPS injection. Urine samples were collected before injection then 12 and 24 hours after LPS administration. Schematic representation of the experimental design (**A**) and ACR levels (**B**) in WT mice injected with either ICOSL protein (blue dots) or BSA (red dots) 24 hours after LPS injection (n=9 per group from two independent experiments). Data are mean±SEM; *P<0.05; Student's two-tailed, unpaired t test (B).



Supplemental Figure 12.

ICOSL treatment had a trend toward lower proteinuria in STZ-induced DN mice.

Urinary albumin excretion (UAE) for 24 hours in ICOSL KO mice treated with either ICOSL protein (blue dots) or BSA (red dots) ($n=6$ per group from one experiment). In brief, male ICOSL KO mice were challenged with STZ to induce diabetes. Two weeks after STZ injection, the mice were divided into 2 groups. Each group ($n=6/\text{group}$) was treated with either mouse ICOSL protein (1 mg/kg, i.p, twice/week) or BSA as a protein control for 4 weeks. For the measurement of urinary albumin excretion, urine samples were collected using metabolic cages at 6 weeks after STZ administration (ICOSL/BSA treatment for 4 weeks). Data are mean $\pm$ SEM; Student's two-tailed, unpaired t test.



Supplemental Figure 13.

The pharmacokinetic (PK) study of ICOSL in mice. *Icosl* KO mice (12-week old female, n=3) were intravenously injected with ICOSL-AF488 conjugate (1 mg/kg body weight). Blood samples (about 30-40 µl) were collected from the retro-orbital plexus at 0.03, 0.5, 1, 3, 6, 9, and 24 hours following ICOSL administration. After spinning down the blood samples at 13,000 rpm for 5 min, sera were obtained and diluted 10-fold with PBS. Standard curve (ranging from 0.156 to 10 µg/ml) was prepared by using serial dilution of the injected ICOSL-AF488 conjugate with PBS containing 10% (v/v) control mouse serum. Fluorescence intensities of standards and diluted sera were measured at 485 nm excitation and 528 nm emission using a fluorescence plate reader (BioTek). The data at each time point were displayed as mean ± SEM and fitted to the equation for two phase decay using Prism version 6.0 (GraphPad).

References

1. Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, Lopez R, McWilliam H, Remmert M, Soding J, et al. Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Mol Syst Biol.* 2011;7(539).